Clincial process to properly prescribe immunotherapy

1st part:

Désirée Larenas-Linnemann, MD, FAAAI, Intl.Dist.FACAAI Clínica de Alergia, Asma e Investigación, Mexico City, Mexico **2nd part:** Noel Rodriguez-Pérez, MD, Intl.Dist.FACAAI, FAAAAI Professor of Pediatrics. State University of Tamaulipas School of Medicine. Matamoros, Mexico

PART I

Correct diagnosis

The clinical process to properly prescribe allergen specific immunotherapy (ASIT) starts with a correct diagnosis of an IgE mediated reaction as the cause of the patient's symptoms. The presence of an IgE mediated hypersensitivity reaction in a patient with clinical signs and symptoms of allergic rhinoconjunctivitis and/or asthma (and/or atopic dermatitis) can be confirmed with in vivo or in vitro testing.¹⁻³

For the in vivo testing the most used method still today is the skin prick test, eventually extended with an intradermal test. However, the latter only adds some diagnostic value in the case of weak diagnostic extracts.⁴⁻⁵Intradermal testing after a negative SPT with a potent extracts, such as a pollen extract in the US, does not add any diagnostic accuracy and thus is not recommended, as the ratio of false positive reactions with an ID test in this scenario is high.¹

Among the in vitro test methods the detection of specific IgE against certain allergens is –at world level- still mostly done with the RAST, Immulite or ImmunoCAP systems. These methods are generally applied for the detection of specific IgE directed against whole allergens. However, newer more specific methods are underway and already on the market in some European countries (very limited in US) that have the potential to detect specific IgE directed against certain epitopes of the allergen. As such, these methods are able to reveal the presence and quantity in patient's serum of highly specific IgE against e.g. Phl p 1, Phl p 2, Phl p 5, etc. instead of only measuring the global IgE against Phleum pratensis. Microarrays are used for these highly specific diagnostic procedures.⁶⁻⁷

However, the sole presence of IgE mediated hypersensitivity is not enough to justify the prescription of ASIT. Two other components are needed to this end:

1) Symptoms of the patient on exposure: a detailed anamnesis is a core tool for the $allergist^{2-3,8}$

2) AND (one of the following)

- Poor response to pharmacotherapy, allergen avoidance, or both;
- Unacceptable adverse effects of medications;
- Wish to reduce or avoid long-term pharmacotherapy and the cost of medication;

- Co-existing allergic rhinitis and asthma;
- Possible prevention of asthma in patients with allergic rhinitis. This indication has been included officially in the WAO statement on sublingual immunotherapy.⁹

Typical indications for SCIT are allergic rhinitis, allergic conjunctivitis, allergic asthma and systemic reactions after exposure to hymenoptera venom. New indications for SCIT are atopic dermatitis in individuals with aeroallergen sensitization and patients who experience recurrent bothersome large local reactions to hymenoptera venom. Though in previous versions of the practice parameters SCIT was not recommended for children below five years of age, in the 3rd update of the PP it is stated that there is no specific upper or lower age limit for initiating allergen immunotherapy.³

ASIT is indicated, and now...

After a correct diagnosis and the decision to start ASIT in a patient, the allergist has to face the next set of challenges:

- 1. Which route: sublingual or subcutaneous?
- 2. Which allergens to include in the immunotherapy? Mono- or multi-allergen ASIT?
- 3. In case of multi-allergen ASIT: which allergens can be mixed together in one vial?

To answer these questions, differences between continents in the practice of immunotherapy must be taken into account. There exist several differences between Europe and North America in the details concerning immunotherapy preparation and application.

- 1. There is no FDA approved extract for sublingual immunotherapy
- 2. In North America the allergist in his office prepares vial for immunotherapy, from which the patient shall receive his/her injections.
- 3. Immunotherapy is generally given with a mixture of several allergens.
- 4. Immunotherapy is given with natural, aqueous extracts. There are neither allergoids nor depot extracts in the US.

Ad 1. Sublingual or subcutaneous immunotherapy

The first decision to be made is about the route of administration. In the US SIT is almost exclusively given as SCIT, because till now there are no FDA approved extracts for SLIT. Even so, a survey among US allergists in 2006 showed that 6% is applying SLIT, using the SCIT extracts.¹⁰ Clinical trials of SLIT have been ongoing the last years in the US. At first they were not too successful.¹¹ but lately good results have been reported, both in adults¹² and in children.¹³

In Mexico and most Latin American (LA) countries both administration routes are available and used by many allergists. (See poster Gómez M, WAC 2011) The sublingual route is especially popular for the application of SIT in (small) children, in patients with particular risk-factors for SCIT, in situations where it is not feasible for the patients to come to the clinic and in some adults who prefer to omit the many office visits for the application of their injections. As such, SLIT and SCIT are considered complementary in LA, as SLIT broadens the market for immunotherapy in general.

Recently, a group of experts cooperated in the development of WAO guidelines on SLIT. Chapter 9 of this document is on the indications for SLIT.¹⁴

Ad 2. Preparation of immunotherapy in the allergist's office

In North America part of the allergist's training is how to prepare allergen immunotherapy.¹⁵ Several points of interest have to be taken into account to adequately prepare a SIT vial, some of which might also apply to the European allergist.

Selection of the allergen(s) to be included in the treatment

Clinically important allergens

Skin prick testing and/or the detection of specific IgE in the patient's serum by Immulite or ImmunoCAP may yield a certain amount of positive results, demonstrating the presence of specific IgE against several allergens. An essential task for the treating allergist is to determine which of these allergens is/are of clinical importance and reduce their number as much as possible. Immunotherapy seems to be more effective, if given with only a few allergens see below.

Cross reactivity

Apart from the selection the allergens for immunotherapy, based on their clinical importance, the number of allergens to be included in the immunotherapy vial can be further reduced taking into account the cross-reactivity between certain allergens, especially those of the same genus or family. Allergens phylogenetically close to one other' in some cases have a high number of similarities. As a consequence, selecting just one of them to cover sensitivities to any other member of the group is enough. This holds especially true for the pollens of certain families of grasses and trees. An excellent review on this subject can be found in two references.¹⁶⁻¹⁷

Proteases

A third rule for the mixing of allergens in one vial has to be taken into account: it is recommended not to put allergens rich in proteases (e.g. molds, cockroach) in the same vial together with allergens with a low concentration of proteases (e.g. grass pollen). Especially for timothy grass pollen it has been shown that mixing with mold extract reduces the detectable amount of allergen after a short while.¹⁸

Immunotherapy with mixes of allergens

A still ongoing point of discussion is the mono- versus multiple allergens SIT, with the former being practiced in Europe and the latter being the way SIT is given in 'the Americas'. Those in favor of the mono-allergen SIT, argue that almost all studies on SIT have been done this an extract of just one allergen and that in most patients treatment with just one -or may be two unrelated- allergen(s) is enough to cover all clinically relevant allergens.

However, those in favor of multiple allergen SIT argument that almost all patients are not only multi-sensitized (=skin prick sensitivity to various non-related allergens), but truly multi-allergic. Moreover, they say, the very first double blind clinical trials showing efficacy for SIT were done with allergen mixes (Johnstone et al 1961¹⁹; Franklin et al, 1967²⁰).

This kind of discussions is interesting, because they show us the multiple faces of allergy: in some areas clear 'hay fever' may exist, as in North and Central Europe, with welldefined pollen seasons of short duration. In this kind of patients surely a mono-allergen SIT is a wonderful solution. However, in other parts of the world -like in some states of the US and many countries of LA- it is rare to find this typical hay fever patient, as pollen is in the air all year long. Moreover, the closer to the equator the higher the concentrations of several species of house dust mite and in the humid sub-tropical areas molds are everywhere. So the allergic patient in these places has a totally different picture of sensitizations and allergies than patients from the aforementioned zones. As a result, immunotherapy probably shall have to be different to be efficacious.

An interesting SLIT study was done in at National Jewish in Denver²¹ rising doubts about the efficacy of multi-mix SLIT: the mono-allergen SLIT performed quite better than the multi-mix SLIT, even though the allergen concentration was kept stable.

Allergen extracts in the US: natural and aqueous

In the US the only extracts on the market are concentrated, aqueous solutions of allergens, meant to be diluted before they can be administered to the patient. A couple of years ago, there were some trials on allergens coupled to adjuvants. One of those molecules is Tolamba, the ragweed pollen major allergen, covalently coupled to Toll-like receptor agonists. However, the FDA has not accepted this kind of immunotherapy yet. For the moment, neither recombinant allergens nor allergoids are used for SIT in the US.

Some European allergen manufacturers have managed to bring their products on the market in some LA countries. Even so the mainstay of SIT in LA is still with aqueous extracts that are mixed in the allergist's office (although there are some exceptions). In Mexico there exist three different categories of commercial allergens: those imported by some Spanish allergen manufacturers and sold as such as end-product; those imported as lyophilized products from raw material providers in the US and sold on the Mexican market unstandardized and the locally manufactured products.²² There is a wide variety in the potency of these three categories²³⁻²⁴

PART II

Allergen dose

Allergists prepare the immunotherapy solution in their offices from concentrated allergen vials. US and LA allergists have taken note of the few dosing studies there exist on Immunotherapy and believe that a certain minimum maintenance dose has to be reached, for IT to be effective.²⁵ Consequently, dosing recommendations exist for projected maintenance doses of each allergen. These doses are meant to be reached in 3-6 months, in a regular regimen. If the patient presents adverse reactions to the applications, it might even take longer before the maintenance dose is reached.

SIT is initiated routinely with approximately a 1/1000th or 1/10.000th of maintenance dose, some times amore diluted extract could be prepared pending on the intensity of ST reaction and slowly build-up with injections once or twice a week. At maintenance, injections are given each 2-4 weeks. The maintenance dose is that considered to provide efficacy without significantly local or systemic adverse reactions. Some patients will not tolerate de projected therapeutic dose and will have to be adjusted to the highest tolerated effective dose of each extract when mixtures are used. This concept does not apply to venom

immunotherapy which the rapeutic dose has to be reached for conferring adequate protection. $^{26}\,$

All dilutions can be calculated with this general formula:^{26a} V1 X C1 = V2 X C2. Were: V1 = Final volume you want to prepare; C1 = Final concentration you want to prepare (w/v, PNU, AU, etc.); V2 (unknown) = Volume of extract (from stock concentrate) you will need for dose required and C2 = Concentration of extract (from concentrate stock) you will use.

Example 1.

Prepare 5 mL (V1) at 1:200 w/v concentration (C1) from a stock concentrate of 1:10 w/v (C2) = V2 (not yet known). We can use either one of the two options of formula composition depending if we are using w/v: $[V2 = (V1/C1) \times C2]$ if we decide no to change fractions for decimals.] or PNU, AU, BAU, etc. $[V2 = (V1 \times C1) / C2]$. Inserting numbers or fractions to the formula, as we need. V1 = 5 mL C1 = 1/200 V2 = ? C2 = 1/10 V1 X C1 = V2 X C2

V2 = (V1/C1) X C2 (5/200) X 10 = 0.25 mL Plus diluent volume (5mL - 0.25 mL) = 4.75 mL

Example 2.

Prepare 5mL (V1) at 2000 AU concentration (C1) from a stock concentration of 10,000AU (C2) = V2 (unknown) V1 = 5 mL C1 = 2000 AU V2 = ? C2 = 10,000 AU V1 X C1 = V2 X C2 V2 = (V1 X C1)/ C2 (5 X 2000)/10,000 = **1.0 mL** Plus diluent volume (5.0 mL - 1.0 mL) = **4.0 mL**

If we need to prepare a mixture of compatible extracts, we have to keep in mind that we have to reach the projected therapeutic dose for each component extract and be aware of the dilution effect between each other.

Example 3.

A total of 10 non-standardized pollen allergens producing positive skin tests have been selected as candidates for IT. The 10 allergens include:

5 trees: box elder, cottonwood, elm, maple, oak 2 grasses: Kentucky blue, timothy 3 weeds: lamb's quarter, pigweed, short ragweed

First, examine the 10 allergens for known patterns of cross- reactivity. Among the 5 trees, box elder and maple belong to the same genus and are highly cross-reactive. The same is

true for the 2 grasses, Kentucky blue and timothy grass. So we will keep only one of each (cottonwood, elm, maple, oak. grasses: timothy. weeds: lamb's quarter, pigweed, short ragweed).

Prepare 5 mL (V1) at 1:200 w/v concentration (C1) from a stock concentrate of 1:10 w/v (C2) = V2 (not yet known). Using w/v [V2 = (V1/C1) X C2.

Inserting numbers or fractions to the formula, as we need.

V1 = 5 mL C1 = 1/200V2 = ? C2 = 1/10V1 X C1 = V2 X C2 V2 = (V1/C1) X C2 (5 /200) X 10 = **0.25 mL X 8 = 2.0 mL** Plus diluent volume (5mL - 0.25 mL) = **3.0 mL**

An individual approach is always recommended. For example, in a patient with four clinically relevant sensitizations e.g. cat, dog, olive pollen and grass pollen, the decision must be based on the allergen which causes:

1. The longest duration of symptoms per year; 2. The most severe symptoms; 3. A major impact on quality of life and 4. Which is more difficult to avoid.

In the previous example, the pet allergens could be avoided, whereas grass pollen is the allergen that has the longest duration of pollen season, however, in some regions the length of pollen season may vary.²⁷.

References

1. Bernstein IL, Li JT, Bernstein DI, Hamilton R, Spector SL, Tan R, et al. Allergy diagnostic testing: an updated practice parameter. Ann Allergy Asthma Immunol. 2008 Mar;100(3 Suppl 3):S1-148.

2. Larenas-Linnemann D, Ortega-Martell JA, Del Rio-Navarro B, Rodriguez-Perez N, Arias-Cruz A, Estrada A, et al. [Mexican clinical practice guidelines of immunotherapy 2011]. Rev Alerg Mex. 2011 Jan-Feb;58(1):3-75.

3. Cox L, Nelson H, Lockey R, Calabria C, Chacko T, Finegold I, et al. Allergen immunotherapy: A practice parameter third update. J Allergy Clin Immunol. 2011 Jan;127(1 Suppl):S1-55.

4. Wood RA, Phipatanakul W, Hamilton RG, Eggleston PA. A comparison of skin prick tests, intradermal skin tests, and RASTs in the diagnosis of cat allergy. J Allergy Clin Immunol. 1999 May;103(5 Pt 1):773-9.

5. Nelson HS, Oppenheimer J, Buchmeier A, Kordash TR, Freshwater LL. An assessment of the role of intradermal skin testing in the diagnosis of clinically relevant allergy to timothy grass. J Allergy Clin Immunol. 1996 Jun;97(6):1193-201.

Sastre J. Molecular diagnosis in allergy. Clin Exp Allergy. 2010 Oct;40(10):1442-60.

7. Hansen KS, Ballmer-Weber BK, Sastre J, Lidholm J, Andersson K, Oberhofer H, et al. Component-resolved in vitro diagnosis of hazelnut allergy in Europe. J Allergy Clin Immunol. 2009 May;123(5):1134-41, 41 e1-3.

8. Walker SM, Durham SR, Till SJ, Roberts G, Corrigan CJ, Leech SC, et al. Immunotherapy for allergic rhinitis. Clin Exp Allergy. 2011 Sep;41(9):1177-200.

9. Canonica GW, Bousquet J, Casale T, Lockey RF, Baena-Cagnani CE, Pawankar R, et al. Sub-lingual immunotherapy: World Allergy Organization Position Paper 2009. Allergy. 2009 Dec;64 Suppl 91:1-59.

10. Coifman RE, Cox LS. 2006 American Academy of Allergy, Asthma & Immunology member immunotherapy practice patterns and concerns. J Allergy Clin Immunol. 2007 Apr;119(4):1012-3.

11. Skoner D, Gentile D, Bush R, Fasano MB, McLaughlin A, Esch RE. Sublingual immunotherapy in patients with allergic rhinoconjunctivitis caused by ragweed pollen. J Allergy Clin Immunol. 2010 Mar;125(3):660-6, 6 e1-6 e4.

12. Nelson HS, Nolte H, Creticos P, Maloney J, Wu J, Bernstein DI. Efficacy and safety of timothy grass allergy immunotherapy tablet treatment in North American adults. J Allergy Clin Immunol. 2011 Jan;127(1):72-80, e1-2.

13. Blaiss M, Maloney J, Nolte H, Gawchik S, Yao R, Skoner DP. Efficacy and safety of timothy grass allergy immunotherapy tablets in North American children and adolescents. J Allergy Clin Immunol. 2011 Jan;127(1):64-71, e1-4.

14. Canonica GW. Sub-lingual immunotherapy: World Allergy Organization position paper 2009. WAO Journal 2010 19jun2010 [cited 2010 19 june]; Available from: http://www.allergychoices.com/school1000162/genie59/images/files/sub_lingual_immunot herapy_world_allergy.11.pdf

15. Nelson M. Allergen Immunotherapy Extract Preparation Guidelines. 2009 [cited 2010 29 sept]; Available from: http://www.aaaai.org/members/resources/allergen_immunotherapy_extract_preparation_gui delines.pdf

16. Weber RW. Cross-reactivity of pollen allergens: impact on allergen immunotherapy. Ann Allergy Asthma Immunol. 2007 Sep;99(3):203-11; quiz 12-3, 31.

17. Weber RW. Guidelines for using pollen cross-reactivity in formulating allergen immunotherapy. J Allergy Clin Immunol. 2008 Jul;122(1):219-21.

18. Grier TJ, LeFevre DM, Duncan EA, Esch RE. Stability of standardized grass, dust mite, cat, and short ragweed allergens after mixing with mold or cockroach extracts. Ann Allergy Asthma Immunol. 2007 Aug;99(2):151-60.

19. Johnstone DE, Crump L. Value of hyposensitization therapy for perennial bronchial asthma in children. Pediatrics. 1961 Jan;27:39-44.

20. Franklin W, Lowell FC. Comparison of two dosages of ragweed extract in the treatment of pollenosis. JAMA. 1967 Sep 18;201(12):915-7.

21. Amar SM, Harbeck RJ, Sills M, Silveira LJ, O'Brien H, Nelson HS. Response to sublingual immunotherapy with grass pollen extract: monotherapy versus combination in a multiallergen extract. J Allergy Clin Immunol. 2009 Jul;124(1):150-6 e1-5.

22. Larenas-Linnemann DL, Cruz AA, Fogelbach GA, del Prado ML. [Allergens used in skin tests in Mexico]. Rev Alerg Mex. 2009 Mar-Apr;56(2):41-7.

23. Larenas-Linnemann D, Matta JJ, Shah-Hosseini K, Michels A, Mosges R. Skin prick test evaluation of Dermatophagoides pteronyssinus diagnostic extracts from Europe, Mexico, and the United States. Ann Allergy Asthma Immunol. 2010 May;104(5):420-5.

24. Larenas-Linnemann D, Cruz AA, Gutierrez IR, Rodriguez P, Shah-Hosseini K, Michels A, et al. European and Mexican vs US diagnostic extracts of Bermuda grass and cat in skin testing. Ann Allergy Asthma Immunol. 2011 May;106(5):421-8.

25. Bernstein DI, Blaiss MS, Cox LS, Finegold I, Lanier BQ, Nelson HS, et al. Current standards and future directions in immunotherapy: perspectives on challenges and opportunities for the allergist. Ann Allergy Asthma Immunol. 2011 Nov;107(5):422-5.

26. Rueff F, Wenderoth A, Przybilla B. Patients still reacting to a sting challenge while receiving conventional Hymenoptera venom immunotherapy are protected by increased venom doses. J Allergy Clin Immunol. 2001 Dec;108(6):1027-32.

26a. Grier TJ. How's My Dosing 2.0: Comparison of 2003-2011 Immunotherapy Practice Parameter Maintenance Dose Recommendations and Updated Math-Free Formulation Tables. Poster presentation at American College of Allergy, Asthma and Immunology annual meeting, Boston MA, November 2011.

27. Zuberbier T, Bachert C, Bousquet PJ, Passalacqua G, Walter Canonica G, Merk H, et al. GA(2) LEN/EAACI pocket guide for allergen-specific immunotherapy for allergic rhinitis and asthma. Allergy. 2010 Dec;65(12):1525-30.